



# Agrin and Synaptic Laminin Are Required to Maintain Adult Neuromuscular Junctions

## Citation

Samuel, Melanie A., Gregorio Valdez, Juan C. Tapia, Jeff W. Lichtman, and Joshua R. Sanes. 2012. Agrin and synaptic laminin are required to maintain adult neuromuscular junctions. PLoS ONE 7(10): e46663.

## Published Version

doi:10.1371/journal.pone.0046663

## Permanent link

<http://nrs.harvard.edu/urn-3:HUL.InstRepos:11729591>

## Terms of Use

This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at <http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA>

## Share Your Story

The Harvard community has made this article openly available.  
Please share how this access benefits you. [Submit a story](#).

[Accessibility](#)

# Agrin and Synaptic Laminin Are Required to Maintain Adult Neuromuscular Junctions

Melanie A. Samuel, Gregorio Valdez<sup>‡a</sup>, Juan C. Tapia<sup>‡b</sup>, Jeff W. Lichtman, Joshua R. Sanes\*

Department of Molecular and Cellular Biology and Center for Brain Science, Harvard University, Cambridge, Massachusetts, United States of America

## Abstract

As synapses form and mature the synaptic partners produce organizing molecules that regulate each other's differentiation and ensure precise apposition of pre- and post-synaptic specializations. At the skeletal neuromuscular junction (NMJ), these molecules include agrin, a nerve-derived organizer of postsynaptic differentiation, and synaptic laminins, muscle-derived organizers of presynaptic differentiation. Both become concentrated in the synaptic cleft as the NMJ develops and are retained in adulthood. Here, we used mutant mice to ask whether these organizers are also required for synaptic maintenance. Deletion of agrin from a subset of adult motor neurons resulted in the loss of acetylcholine receptors and other components of the postsynaptic apparatus and synaptic cleft. Nerve terminals also atrophied and eventually withdrew from muscle fibers. On the other hand, mice lacking the presynaptic organizer laminin- $\alpha 4$  retained most of the synaptic cleft components but exhibited synaptic alterations reminiscent of those observed in aged animals. Although we detected no marked decrease in laminin or agrin levels at aged NMJs, we observed alterations in the distribution and organization of these synaptic cleft components suggesting that such changes could contribute to age-related synaptic disassembly. Together, these results demonstrate that pre- and post-synaptic organizers actively function to maintain the structure and function of adult NMJs.

**Citation:** Samuel MA, Valdez G, Tapia JC, Lichtman JW, Sanes JR (2012) Agrin and Synaptic Laminin Are Required to Maintain Adult Neuromuscular Junctions. PLoS ONE 7(10): e46663. doi:10.1371/journal.pone.0046663

**Editor:** Brian D. McCabe, Columbia University, United States of America

**Received:** August 2, 2012; **Accepted:** September 4, 2012; **Published:** October 3, 2012

**Copyright:** © 2012 Samuel et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** This study was funded by grants R01AG32322 and R01NS59853 from the NIH. The funder had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

\* E-mail: sanesj@mcb.harvard.edu

<sup>‡a</sup> Current address: Virginia Tech Carilion Research Institute, Roanoke, Virginia, United States of America

<sup>‡b</sup> Current address: Department of Neuroscience, Columbia University, New York, New York, United States of America

## Introduction

At the mammalian skeletal neuromuscular junction (NMJ), pre- and post-synaptic structures are highly differentiated and precisely aligned. This synapse forms in embryos, matures during an early postnatal period, and is remarkably stable in young adult mice [1–3]. Later in life, however, maintenance of the NMJ is compromised, and several structural alterations occur. By two years of age, NMJs in most muscles show marked dystrophies, including nerve terminal sprouting, fragmentation, and denervation [4–9]. Related patterns of progressive NMJ disassembly have also been documented in some diseases of the motor unit, including myasthenia gravis and amyotrophic lateral sclerosis [9–13]. These alterations presumably result from changes in the level or distribution of molecules that maintain synaptic structure and function in young adults. To date, however, the identity of such factors has remained obscure.

In seeking such factors, we considered molecules called synaptic organizers, which are already known to regulate the formation and maturation of the NMJ [14–16]. Several synaptic organizers are secreted by motor axon terminals or muscle fibers and are stably maintained in the basal lamina that runs through the synaptic cleft between the nerve terminal and postsynaptic membranes [17–21]. Of these, the two best characterized are agrin and the synaptic laminins.

Agrin is a nerve-derived proteoglycan that is secreted by motor axons and promotes aggregation of acetylcholine receptors (AChRs) in the postsynaptic membrane. Muscle cells also express agrin, but only motor neurons produce an alternatively spliced isoform called “z-agrin” that activates a postsynaptic receptor complex consisting of LRP4 and the receptor tyrosine kinase MuSK [22–25]. During the initial phases of development, z-agrin activates MuSK/LRP4 to both stabilize aneurally-formed clusters of AChRs and induce formation of new clusters in the muscle membrane [26–30]. In the absence of all agrin isoforms or only z-agrin, postsynaptic differentiation of the NMJ does not proceed, and animals die shortly after birth [18,19]. Presynaptic abnormalities are also evident in agrin mutant mice, likely because disruption of postsynaptic development prevents retrograde signaling from muscle to nerve.

In contrast to nerve-derived agrin, laminins are synthesized by muscles and promote presynaptic differentiation. Laminins are present in all basal laminas (BLs) and are heterotrimers of  $\alpha$ ,  $\beta$ , and  $\gamma$  subunits drawn from a total of 5 $\alpha$ , 3 $\beta$ , and 3 $\gamma$  isoforms [31]. There are specific laminin heterotrimers in the synaptic cleft, laminins 221 ( $\alpha 2\beta 2\gamma 1$ ), 421 ( $\alpha 4\beta 2\gamma 1$ ), and 521 ( $\alpha 5\beta 2\gamma 1$ ); extrasynaptic BL contains primarily laminin 211 ( $\alpha 2\beta 1\gamma 1$ ) [32]. In laminin- $\beta 2$  mutant mice all synaptic laminin heterotrimers are depleted, the NMJ fails to mature and animals die 2–4 weeks after birth due to a lack of proper neuromuscular

transmission [17,33]. Mice lacking laminin- $\alpha 4$  (*lama4*<sup>-/-</sup>) are viable but lack precise apposition of presynaptic neurotransmitter release sites (active zones) to postsynaptic depressions called junctional folds [20]. Mice lacking both laminins- $\alpha 4$  and - $\alpha 5$  also show presynaptic abnormalities as well as delayed postsynaptic maturation [34].

Here, we asked whether agrin and synaptic laminins are required only for synaptic development and maturation, or whether their continuous presence is also required for synaptic maintenance in adult NMJs. To bypass the neonatal lethality of agrin mutants, we used a conditional allele (*agrn*<sup>flax</sup>, ref [35]) to selectively delete agrin from a marked subset of motoneurons in adults. Agrin deletion resulted in the depletion of AChR receptors, dispersion of basal lamina components, NMJ dystrophies, and eventually nerve retraction. Lacking a conditional laminin- $\beta 2$  mutant, we used *lama4* mutants, which show subtle developmental effects but are otherwise normal and fertile [20]. By 6 months of age, NMJs lacking laminin- $\alpha 4$  exhibited severe structural defects. Abnormalities in both *agrn*<sup>flax</sup> and *lama4* mutants were distinct from each other, but resembled in several respects those seen in NMJs of aged wild-type mice. Moreover, the synaptic distribution of agrin and synaptic laminins changed with age. Together, these results suggest that maintenance of the NMJ requires the continuous presence of synaptic organizers and that alterations in the integrity of the synaptic basal lamina may contribute to age-related synaptic disassembly.

## Results

### Agrin Depletion by Conditional Deletion in Adults

Mice lacking all agrin isoforms or only  $\alpha$ -agrin die soon after birth due to severe neuromuscular defects [18,19]. To bypass lethality, we used agrin conditional mutants (*agrn*<sup>flax</sup>) in which the genomic sequence from exon 6 to intron 33 is flanked by loxp sites [35]. For deletion in adults, we used a transgenic line termed SLICKA in which a tamoxifen-activated derivative of Cre recombinase (Cre-estrogen receptor fusion or Cre-er) and YFP are co-expressed in a subset of motor neurons under the control of regulatory elements from the *Thy1* gene [36]. Administration of tamoxifen deletes agrin from only those motoneurons that are YFP positive (YFP+) (**Fig. 1A, B**). In these experiments, we administered 8 doses of tamoxifen over 2 weeks [36] to maximize the fraction of YFP+ motoneurons in which agrin deletion occurred. Thus, many YFP positive motoneurons are agrin negative, and all YFP negative (YFP-) motor neurons are agrin positive, allowing comparison of wild-type and agrin-deficient NMJs in the same muscle. In the experiments presented here, we administered tamoxifen to 2–3 month old mice; to facilitate agrin deletion, experiments were performed with null/conditional trans-heterozygotes (*agrn*<sup>-/flax</sup>).

In initial studies, we assessed levels of agrin at the NMJ by immunohistochemistry 1 to 3 months post-tamoxifen injection. As expected, agrin was concentrated at NMJs in control muscle (**Fig. 1C**, left panel) and at YFP- NMJs in mutants (data not shown). In contrast agrin levels decreased at most YFP+ NMJs in mutants. Modest decreases could be observed as early as one month, but complete loss did not occur until 2 to 3 months after tamoxifen was administered (**Fig. 1C**, right panel). Nerve terminals were also altered; this result is discussed below.

### Agrin is Required for Maintenance of the Postsynaptic Apparatus

The best-studied role of agrin is as a factor that promotes the stabilization of AChR clusters. To ask whether agrin is required

for maintenance of these aggregates in adult muscle, we stained triangularis sterni muscle with fluorophore tagged  $\alpha$ -bungarotoxin (f-BTX), which binds tightly and specifically to AChRs. In control heterozygous animals (*agrn*<sup>+/-flax</sup>; SLICKA<sup>+</sup>) AChRs remained normally clustered 3 months after tamoxifen administration, and terminal branches of individual axons exactly opposed postsynaptic sites (**Fig. 2A**, left panel). In contrast, AChR receptor clusters were undetectable at a large number of YFP+ axon terminals in agrin mutants (**Fig. 2A**, right panel, arrows). Thus agrin is required for maintenance of AChR clusters at adult NMJs.

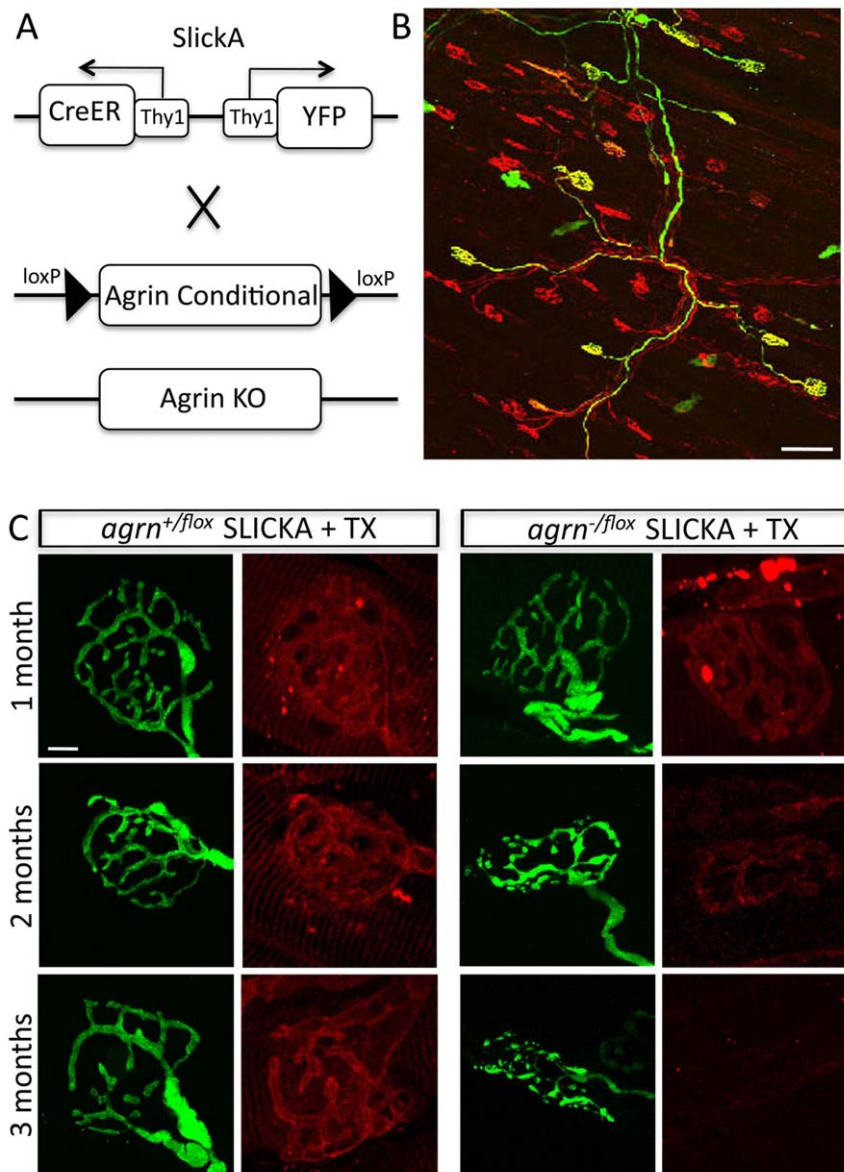
During development, agrin is required for most if not all aspects of postsynaptic maturation [18,19]. To ask whether agrin is also required for the maintenance of the postsynaptic apparatus, we immunostained mutant muscles with antibodies to two components of the synaptic basal lamina (laminin- $\alpha 4$  and - $\alpha 5$ ), a component of the postsynaptic membrane ( $\beta$ -dystroglycan) and a component of the subsynaptic cytoskeleton (rapsyn). We focused on synapses that displayed structural defects as these show the highest level of agrin depletion (see below). Levels of laminin- $\alpha 4$  and - $\alpha 5$ , both of which are concentrated in synaptic basal lamina at control NMJs, were markedly reduced in affected synapses (**Fig. 2C, D**). Likewise, levels of  $\beta$ -dystroglycan, a laminin receptor, and of rapsyn, a cytoskeletal protein that binds AChRs, were depleted (**Fig. 2E, F**). We also stained muscles with antibodies to MuSK, but staining was unreliable in control muscles, so we cannot draw conclusions about its level or distribution in mutants. Interestingly, there appeared to be a clear correspondence between protein levels and AChRs intensity, such that the intensity of AChRs at a particular NMJ was correlated with that of the other component examined: high, mid, and low levels of AChRs were associated with high, mid, and low levels of the other synaptic proteins (data not shown).

### Agrin Loss Disrupts Nerve Terminal Organization

To characterize presynaptic alterations at NMJs depleted of agrin, individual YFP+ nerve terminals from control and agrin mutant animals were imaged at a high magnification together with f-BTX at 3 months after tamoxifen injection (**Fig. 3A, B**). Some YFP+ nerve terminals in mutants were apposed to postsynaptic sites containing normal levels of AChRs (**Fig. 3A**). The majority of these terminals were structurally normal, presumably because the *agrn* gene had not been disrupted in the parent neuron. Consistent with this idea, presynaptic alterations seen at 3 months after deletion occurred predominantly at NMJs from which agrin was depleted (**Fig. 3C**).

In contrast, terminals at NMJs with decreased AChR levels displayed presynaptic structural defects that ranged in severity from little alteration to severe disruption (**Fig. 3A**). Affected mutant terminals differed from those in control animals in several ways. In some cases, axon terminals displayed thinning and dystrophy, while in others the terminals showed fragmentation or swelling (**Fig. 3B**, arrows). Sprouting from a terminal occurred in rare cases. Together, these data suggest that agrin is required for maintaining AChR and axonal terminal patterning in adults.

In comparing pre- to postsynaptic alterations, we noted some cases in which relatively intact presynaptic structures were apposed to defective postsynaptic structures, but very few cases in which structurally abnormal nerve terminals apposed normal appearing postsynaptic structures. In addition, the majority of AChR aggregates in these muscles were unopposed by a YFP+ axon presumably because they were innervated by a YFP- axon. However, it was also possible that some AChR clusters had



**Figure 1. Agrin is depleted from NMJs following conditional deletion in adults.** A. Schematic of the method used to delete agrin from subsets of motoneurons. In the SLICKA transgene, separate Thy1 regulatory elements drive expression of CreER and YFP. Following administration of tamoxifen, CreER is activated and the agrin gene is deleted in many YFP+ motoneurons. B. A YFP+ motor axon (green) in a triangularis sterni muscle. Unlabeled axons are stained with neurofilament and syntrophin 2 (red) to mark all nerve terminals. Scale bar = 100  $\mu$ m. Explain the red labeling of motor axons \*\*\*SLICK negative = Neurofilament/Syn???. C. Agrin at NMJs in control (*agrn*<sup>+/flox</sup>; SLICKA) and mutant (*agrn*<sup>-flox</sup>; SLICKA) muscle at the indicated times after tamoxifen (TX) administration. Agrin levels are decreased slightly 1 month after TX and markedly by 3 months. Scale bar = 10  $\mu$ m. TX = tamoxifen.  
doi:10.1371/journal.pone.0046663.g001

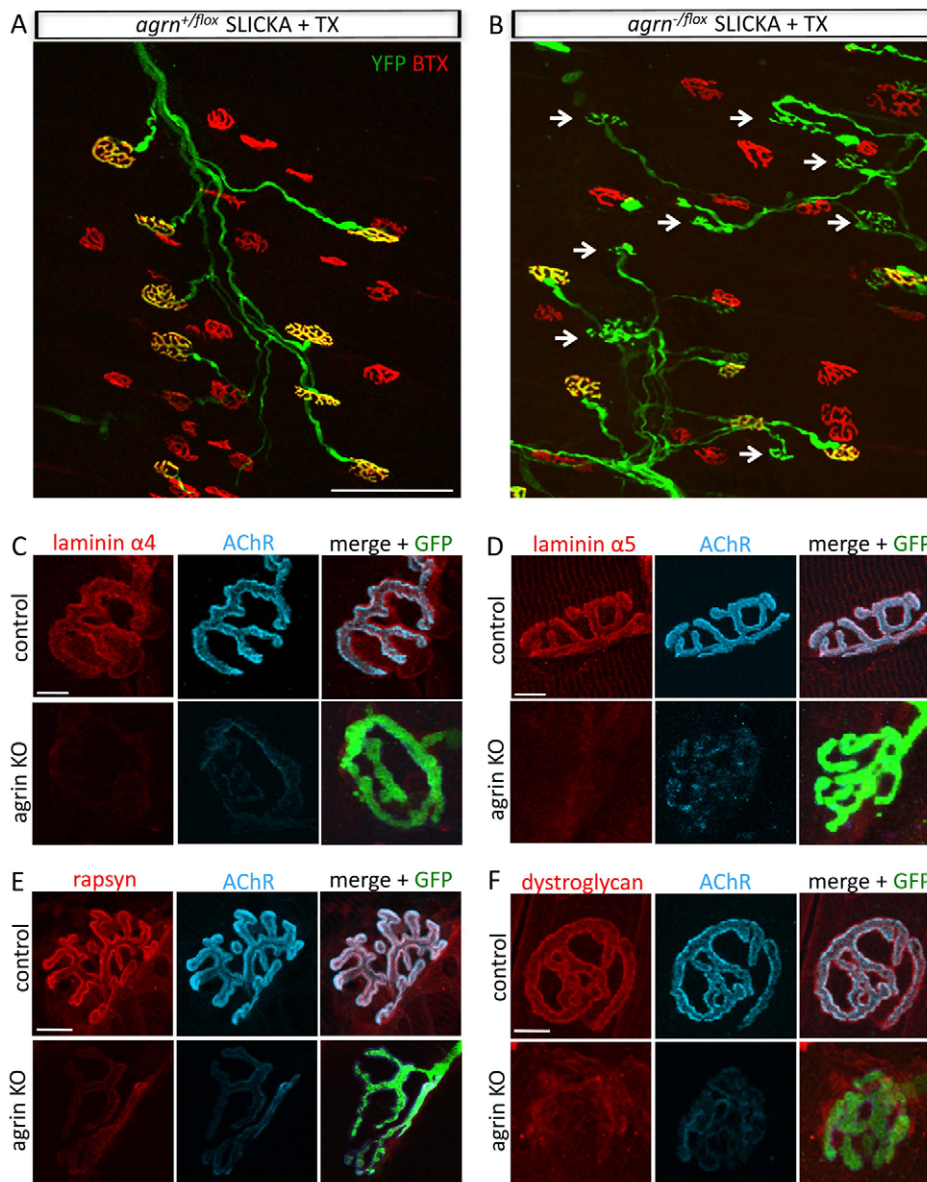
become denervated – that is, they had lost YFP+ innervation. To test this possibility, we stained all axons with antibodies to neurofilaments and found no endplates that lacked innervation (data not shown). Together, these results suggest that postsynaptic alterations precede presynaptic alterations.

To determine the sources of variability among affected nerve terminals, we performed three additional studies. First, we determined the fraction of YFP+ terminals with structural alterations at monthly intervals following tamoxifen administration. The percent of YFP+ terminals exhibiting one or more of the alterations described above increased from ~12% at 1 month post deletion when agrin levels are fairly normal, to ~75% at 3 months

when agrin is largely depleted (**Fig. 3D**). As noted above, most of the YFP+ terminals that appeared normal at 3 months apposed agrin-positive postsynaptic sites, and therefore likely arose from motor neurons in which *agrn* had not been deleted.

Second, we examined several muscles that differ in position and fiber type composition. Changes in AChR clustering and nerve terminal architecture similar to those documented in Fig. 3 for triangularis sterni were observed in hindlimb, trunk and facial muscles (e.g. gastrocnemius, tibialis anterior, and levator auris longus, and interscutularis; data not shown). Thus, most or all NMJs depend on agrin for their maintenance.

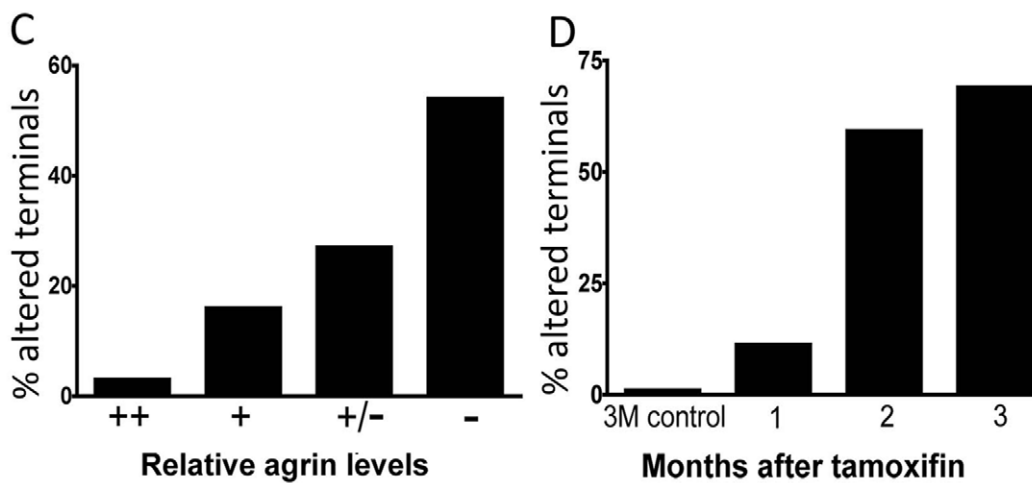
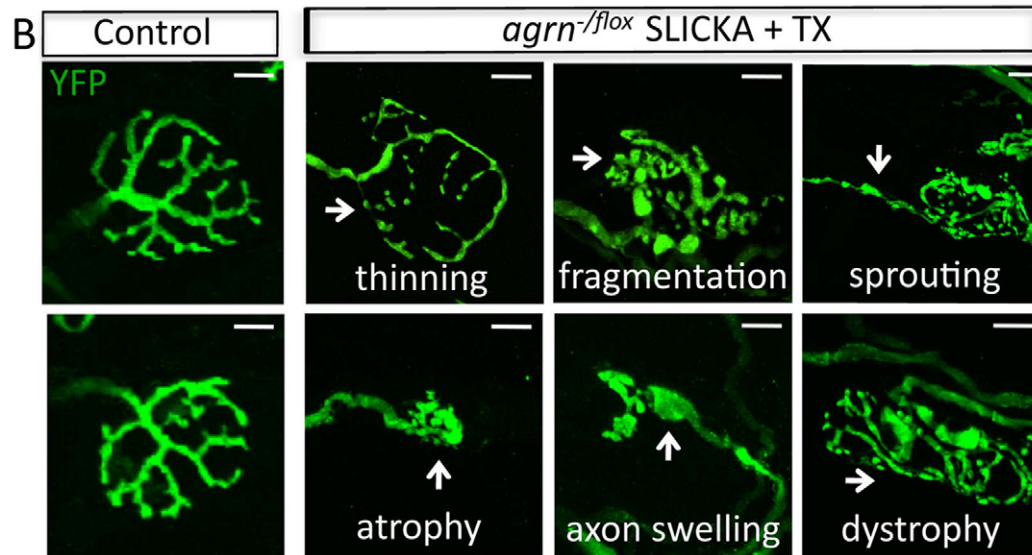
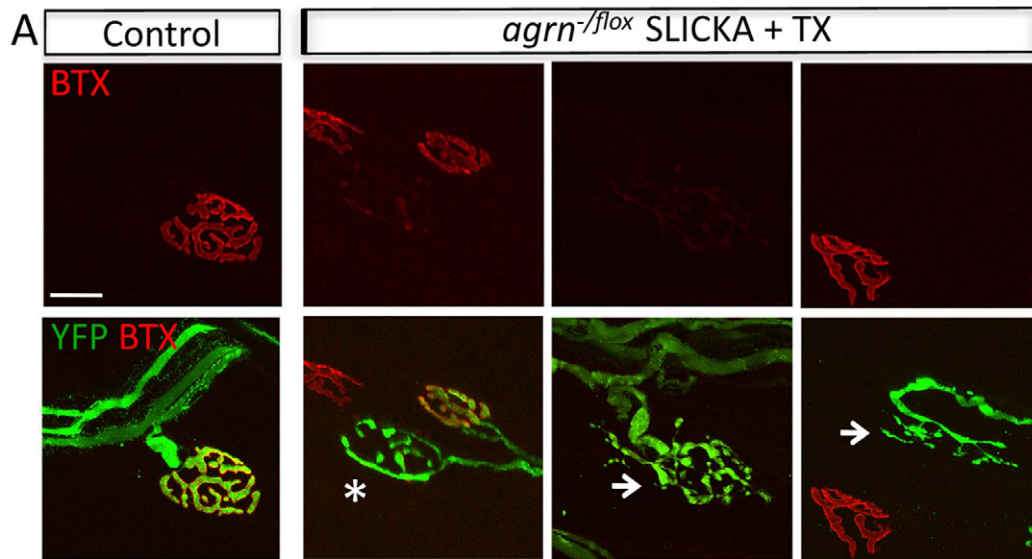




**Figure 2. Agrin loss leads to disassembly of the postsynaptic apparatus.** A–B. Triangularis sterni muscle from control (A) and conditional agrin knockout (B) animals 3 months after tamoxifen (TX) administration. TX administration to heterozygotes does not disrupt apposition of axons (green) to AChR-rich postsynaptic membrane (red), but many axons appose AChR-poor postsynaptic structure (arrows) in the mutant. Scale bar = 100  $\mu$ m. TX = tamoxifen C–F Muscles from agrin conditional knockouts and controls 3 months after TX stained for laminin  $\alpha$ 4 (C), laminin  $\alpha$ 5 (D), rapsyn (E), and dystroglycan (F). In each case, mutant NMJs show markedly decreased levels of immunoreactivity. Scale bar = 10  $\mu$ m. doi:10.1371/journal.pone.0046663.g002

Third, we examined the variability in structural alterations among nerve terminals arising from a single motor axon. Because multiple YFP+ axons were present in most muscles, it was not feasible to reconstruct entire motor units, so we examined pairs of terminals in sparsely labeled regions that we knew arose from a single axon because we could trace them back to a common branch point. In nearly all cases the terminal structure of each member of a pair was similarly affected (**Fig. 4A**) and in <5% of pairs was one NMJ normal and the other clearly abnormal (**Fig. 4B**). These data support the idea that structural defects reflect the level of agrin within a motor unit and suggest that variation among NMJs may result from differences in the rate at which agrin is lost following deletion.

Finally, we extended our analysis to 5–6 months after tamoxifen injection to assess the fate of motor axons lacking agrin. By this time, pretzel shaped presynaptic nerve terminals had nearly all disappeared (**Fig. 5A**). The terminal structures that remained showed severely disrupted morphology with markedly simplified arbors; adjacent regions of muscle fibers were AChR-poor (**Fig. 5B**). In many cases, axons appeared to have withdrawn from synaptic sites, leaving a single fine-tipped or bulbous ending (**Fig. 5C**). Branching of YFP+ axons in mutants was much less complex than in control animals, leading to decreased size of the motor unit (**Fig. 5A** and data not shown). Moreover, the number of YFP+ axons per triangularis muscle was lower at 6 months after tamoxifen (1–3 per muscle) than at 1 or 3 months (4–10 per muscle). Thus, having lost their NMJs, the tamoxifen-deficient



**Figure 3. Agrin loss disrupts nerve terminal organization.** A. Variation in nerve terminal morphology at AChR-poor NMJs in agrin mutants. Some terminals appear relatively normal (\*) while others show marked dystrophies (arrows). Scale bar = 20  $\mu$ m. B. Range of presynaptic defects in agrin mutant nerve terminals (arrows). YFP, green. C–D. The terminals that are affected show low levels of agrin relative to those that are unaffected (C), and the percentage of terminals that are altered increases over time (D). n = at least 4 samples per group. doi:10.1371/journal.pone.0046663.g003

axons exhibit a “dying back” neuropathy typical of ALS and other axonal degeneration diseases [10].

### Presynaptic Alterations in NMJs of Laminin- $\alpha$ 4 Mutants

To determine the role of muscle-derived organizers in synaptic maintenance we examined the role of laminin- $\alpha$ 4 in adult NMJs. In previous studies, we showed that young adult *lama4*<sup>−/−</sup> mice are viable and fertile but have small NMJs in which active zones are misaligned with postsynaptic junctional folds [20]. Here, we asked whether additional alterations occurred at later ages. We stained muscles from control and *lama4*<sup>−/−</sup> mice with anti-neurofilaments to label axons, anti-synaptotagmin-2 to visualize nerve terminals, f-BTX to label AChRs, and anti-laminin- $\alpha$ 4 to confirm deletion. At 40 days of age (sexual maturity), *lama4*<sup>−/−</sup> NMJs were smaller than those controls, but appeared largely normal by light microscopic criteria (data not shown; see ref [20]). At 6 months of age, however, nerve terminals were swollen and/or partially withdrawn from ~50% of NMJs in *lama4*<sup>−/−</sup> mice (Fig. 6). In addition, whereas control NMJs are comprised of long branches, many mutant NMJs exhibited a fragmented appearance, with both nerve terminals and the AChR-rich postsynaptic apparatus broken into round or ovoid plaques.

To ask whether loss of laminin- $\alpha$ 4 affected the molecular architecture as well as the geometry of the postsynaptic apparatus, we stained with a panel of antibodies to synaptic cleft and postsynaptic membrane components. We observed no marked difference between mutant and control NMJs in the levels of laminin-B2, laminin- $\alpha$ 5, agrin,  $\beta$ -dystroglycan or rapsyn (Fig. 7 and data not shown). The results are consistent with the idea that laminin- $\alpha$ 4 exerts its effects directly on nerve terminals rather than

regulating postsynaptic components directly and nerve terminals secondarily.

### Premature Aging of NMJs in Laminin- $\alpha$ 4 Mutants

The structural alterations observed in motor nerve terminals of 6 month-old *lama4* mutants resembled those observed in NMJs of 1.5–2 year old wild-type mice [8]. To quantify these alterations, we crossed *lama4* mutants to Thy1-YFP16 mice, which express YFP in all motoneurons [37]. We then determined the fraction of NMJs in diaphragm muscle that exhibited each of 6 features characteristic of aging NMJs: fragmentation of branches into small islands, partial or complete denervation of the postsynaptic membrane, terminal sprouting, axonal dystrophy and multiple innervation of individual synaptic sites (Fig. 8A). Results from control diaphragms were similar to those documented recently for hindlimb muscles [8]: the incidence of alterations was extremely low at 1 and 6 months of age, and then increased markedly between 1 and 2 years of age. In striking contrast, alterations were abundant in *lama4*<sup>−/−</sup> muscle at 6 months of age and increased further by 10 months of age (Fig. 8B). By these criteria, NMJs accumulated age-related alterations 6–10 months earlier in absence of laminin- $\alpha$ 4 than in its presence.

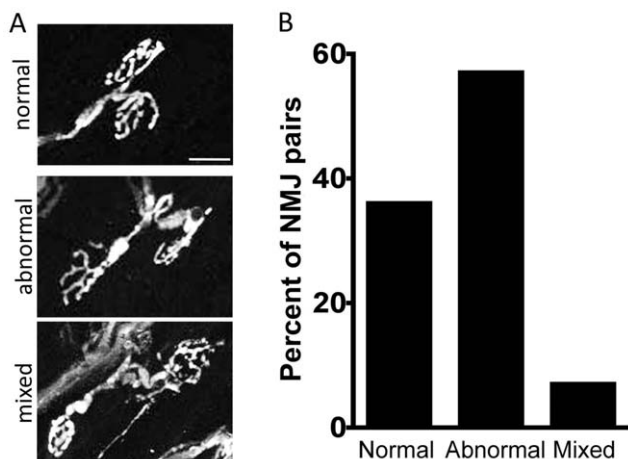
### Alterations in the Synaptic Cleft of Aged NMJs

Since deletion of agrin and laminin- $\alpha$ 4 cause premature disassembly of the NMJ, we asked whether alterations in these molecules occur in aging NMJs. To test for transcriptional changes, we carried out quantitative PCR on samples from old and young mice using primers specific to different agrin isoforms and laminin- $\alpha$ 4, - $\alpha$ 5 and - $\beta$ 2 chains. We assayed agrin and laminin RNA levels in spinal cord and muscle, respectively, because the bioactive z-agrin in the synaptic cleft is derived exclusively from motoneurons whereas the synaptic laminins are synthesized in muscle fibers [38,39]. The abundance of agrin mRNA (assayed with primers that amplify all isoforms) and z-agrin (assayed with exon-specific primers) did not differ detectably between spinal cords from young and old mice (Fig. 9A). Likewise, abundance of mRNAs encoding the three laminin chains changed little if at all in old muscles (Fig. 9B).

We next asked whether aging affected levels or distribution of these molecules at the NMJ. Young adult and aged whole triangularis muscles were stained with antibodies against agrin and the three synapse localized laminin chains. We detected no difference in levels of agrin or laminins between young adult and old muscles, but their distribution was markedly altered. Whereas all four proteins were precisely apposed to regions of high AChR density in young adults, some synaptic regions were depleted of these proteins in old mice, and immunoreactivity extended into perisynaptic areas (Fig. 9C–F).

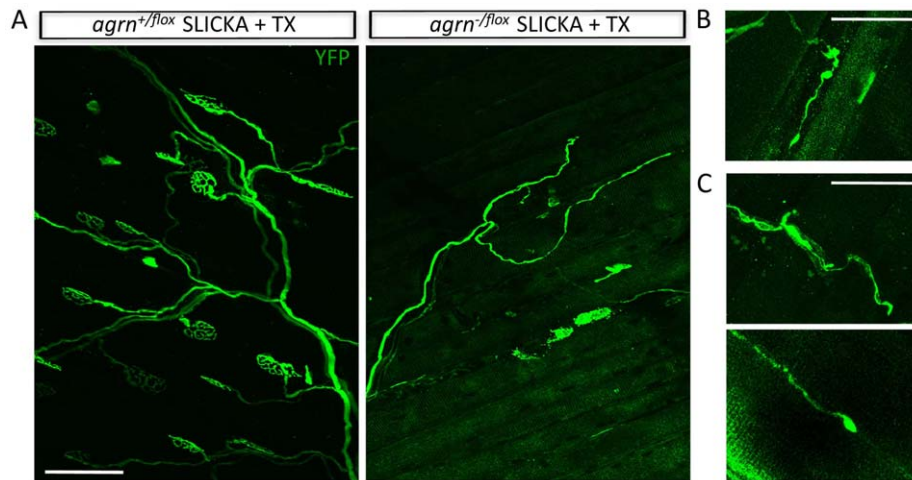
### Discussion

A key feature of the adult NMJ is that it is remarkably stable under ordinary circumstances [1–3] yet capable of remodeling in response to altered circumstances such as increased load or denervation of nearby fibers [40–43]. This combination of stability and malleability implies that synaptic maintenance is controlled actively, yet little is known about the underlying molecular



**Figure 4. NMJs From the same motor neuron are similarly affected following conditional agrin deletion.** A. Three pairs of nerve terminals from triangularis muscles of conditional agrin mutant 3 months after tamoxifen administration. In each case, both terminals arise from the same axon. Both are unaffected in one pair and both are affected in a second pair; in the third pair, one terminal is normal and the other is dystrophic. Scale bar = 20  $\mu$ m. B. Quantification of nerve terminal pairs. n = 100 total pairs. doi:10.1371/journal.pone.0046663.g004





**Figure 5. Nerve degeneration and death results from long-term agrin deletion.** A. YFP labeled axons in the triangularis of control and agrin mutant mice 6 months after tamoxifen administration. Axons in the mutant show less branching than those in controls. Scale bar = 100  $\mu$ m. B–C. Mutant nerve terminals show little to no presynaptic specialization (B) but rather often end in a single axon terminal bulb (C). Scale bar = 50  $\mu$ m. YFP, green.

doi:10.1371/journal.pone.0046663.g005

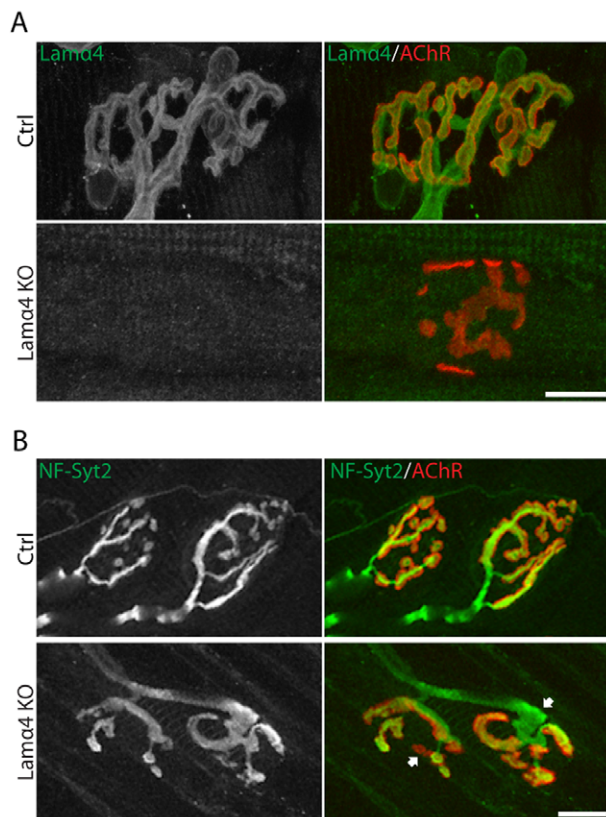
regulatory mechanisms. In this study, we used genetic methods to show that two synaptic organizing molecules known to regulate development of the NMJ, agrin and laminin- $\alpha$ 4, are also required

for synaptic maintenance in adults. We documented progressive presynaptic alterations in mice lacking laminin- $\alpha$ 4 (*lama4*<sup>-/-</sup>) and synaptic disassembly following deletion of agrin in adults (*agrn*<sup>flox/+</sup>;SLICKA). Alterations observed in the two mutants are related but distinct, and both resemble some of those seen during normal aging and in diseases of the motor unit [9–13]. We conclude that multiple synaptic organizers cooperate not only to form the NMJ during embryonic and early postembryonic life but also to maintain it in adults. Our results also raise the possibility that alterations in these or other organizing molecules are involved in age-dependent disassembly of the NMJ.

### Agrin is Required for Pre- and Postsynaptic Maintenance

By using a conditional agrin mutant (*agrn*<sup>flox</sup>), we could allow synapses to develop normally and then assess the effect of agrin loss in adulthood. A role for agrin in synaptic maintenance was not unexpected given its critical role in development [18,19]. However, in the absence of innervation, and thus z-agrin, AChR aggregates form and other components of the postsynaptic membrane and synaptic cleft accumulate both in vitro and in vivo [26,27,44]. In contrast, the agrin co-receptor, MuSK is required for AChR aggregation under both normal and aneural conditions [22,26,27]. These and other results have led to a model in which basal levels of MuSK signaling initiate synapse formation, with agrin increasing MuSK activity to stabilize postsynaptic structures and induce formation of additional ones [15,16]. Consistent with this idea, modest increases in MuSK levels in muscle, achieved transgenically, rescue the lethality of agrin mutants [45]. In view of these results, it was an open question whether agrin would be required to maintain appropriate AChR clustering in mature nerve terminals where other synaptic organizers remain.

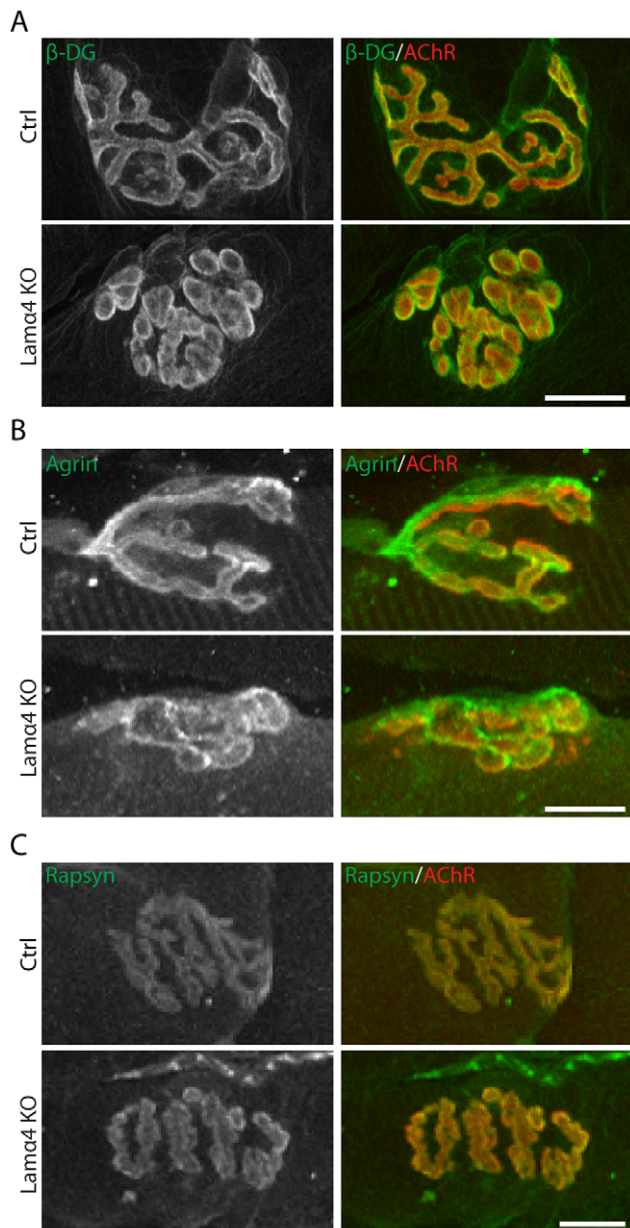
Agrin depletion induced dramatic alterations at the NMJ: (1) AChR aggregates were lost from the postsynaptic membrane; (2) other components of the postsynaptic membrane ( $\beta$ -dystroglycan), subsynaptic cytoskeleton (rapsyn), and synaptic cleft (laminins- $\alpha$ 4 and - $\alpha$ 5) were depleted; and (3) nerve terminals remodeled, became dystrophic and eventually withdrew from the end-plate. Why does loss of agrin lead to so many structural alterations at the NMJ? The best-characterized role of agrin is to induce and stabilize AChR clusters. Other alterations could either reflect direct roles for agrin in



**Figure 6. Abnormal NMJs in laminin- $\alpha$ 4 knockout mice.** A–B. Laminin- $\alpha$ 4 is completely absent in the NMJs from 6 month old *lama4*<sup>-/-</sup> mice (A), and at this time point the NMJ has begun to atrophy (B). There are clear signs of axonal swellings (arrow) and denervation (star) in control 6 months old mice. Scale bar = 10  $\mu$ m.

doi:10.1371/journal.pone.0046663.g006





**Figure 7. Deletion of laminin- $\alpha$ 4 does not alter the levels or localization of other synaptic components.** Distribution and levels of  $\beta$ -dystroglycan (A), agrin (B) and rapsyn (C) in 6 month old control and *lama4*<sup>-/-</sup> NMJs. All three proteins remain concentrated in NMJs lacking laminin- $\alpha$ 4. Scale bar = 10  $\mu$ m. doi:10.1371/journal.pone.0046663.g007

many aspects of synaptic maintenance, or secondary effects of AChR loss. In favor of the latter possibility is the observation that postsynaptic defects appear to precede presynaptic ones: presynaptic alterations are found only at defective postsynaptic structures whereas postsynaptic alterations often occur at synapses with structurally normal nerve terminals. An attractive model is that dispersion of AChRs leads to rapid loss of other components, including presynaptic organizers such as  $\beta$ 2 laminins and LRP4 [46], which in turn leads more slowly to presynaptic changes. It is also possible, however, that agrin has multiple direct effects. In favor of this alternative, agrin treatment can cluster some synaptic proteins when applied to muscle cells that lack AChRs [47] and it can act directly on

axons to affect growth, differentiation and adhesion [48–50]. Indeed, agrin-MuSK signaling is thought to encode an axon growth ‘stop signal’ as mutating either protein leads to exuberant axon sprouting [18,19,51]. It was perhaps surprising, then, that we only rarely detected axon sprouting in adult conditional agrin mutants. This may reflect either inherent differences in adult axon growth capacity or indicate that some functions of agrin are restricted to specific time windows.

The slow pace of change following *agm* deletion presumably reflects the slow loss of z-agrin from the synaptic cleft. Although we have no data on the rate at which *agm* mRNA and protein levels decline within motoneurons following gene inactivation, our results are consistent with the idea that agrin turns over slowly, as has been shown for other basal lamina components in many tissues, and for recombinant agrin injected into muscles [52]. Slow turnover could be an important factor in explaining the limited stability of postsynaptic sites following denervation. Although postsynaptic structures are maintained for weeks following denervation, AChR density subsequently declines and the postsynaptic apparatus disassembles in 2–3 months [53], a time consistent with the disappearance of agrin following *agm* deletion.

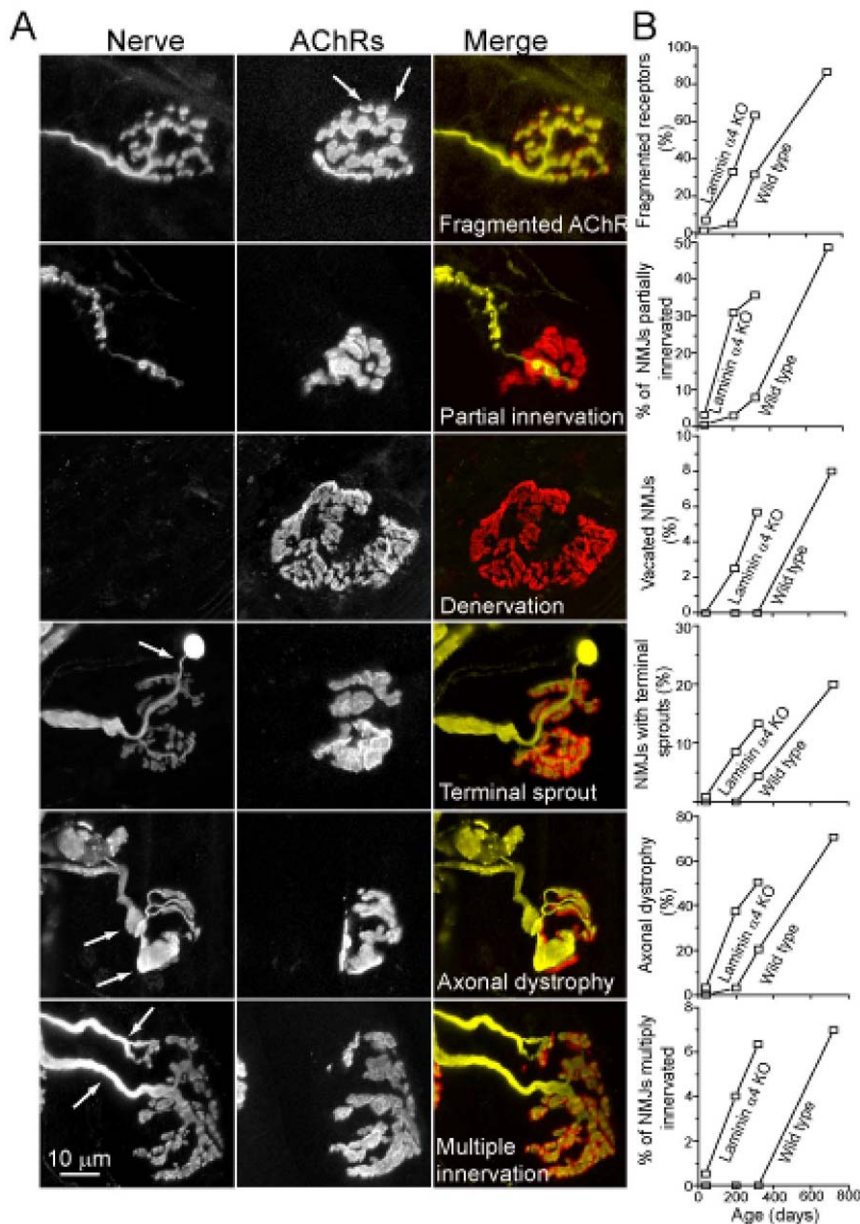
### Laminin- $\alpha$ 4 is Required for Maintenance of Nerve Terminals

To assess roles of synaptic laminins in maintenance of the NMJ, we examined mice lacking laminin- $\alpha$ 4. Five laminin subunits are present in the basal lamina of the synaptic cleft – $\alpha$ 2,  $\alpha$ 4,  $\alpha$ 5 and  $\beta$ 2 and  $\gamma$ 1– of which  $\alpha$ 4,  $\alpha$ 5 and  $\beta$ 2 are present at several-fold higher levels synaptically than extrasynaptically. The rationale for analyzing  $\alpha$ 4 was as follows: lack of either broadly distributed subunit,  $\alpha$ 2 or  $\gamma$ 1, has devastating effects on muscle integrity. Lack of  $\beta$ 2 severely perturbs presynaptic development, but the null mutant is lethal [17] and no conditional mutant has been reported. Lack of  $\alpha$ 5 delays maturation of the NMJ but does not affect its adult form [34]. In contrast, laminin- $\alpha$ 4 is required for precise apposition of pre- and postsynaptic structures at the NMJs in young adults, but does not detectably compromise the topology or molecular architecture of the synapse, and *lama4*<sup>-/-</sup> mice are viable and fertile. We were therefore able to ask whether laminin- $\alpha$ 4 is required for long-term maintenance of the NMJ as well as its earlier maturation.

In contrast to the complete synaptic disassembly that occurred following loss of neural agrin in adults, effects of deleting laminin- $\alpha$ 4 were largely confined to presynaptic structures. Moreover, loss of laminin- $\alpha$ 4 had no obvious effect on the molecular architecture of the synaptic cleft or postsynaptic membrane, either in young adults [20] or in 6–12-month old mice. Together, these data suggest that laminin- $\alpha$ 4 acts directly on nerve terminals rather than indirectly via postsynaptic disruption or coordinate changes in the synaptic cleft. Laminin- $\alpha$ 4 is concentrated in punctae within the synaptic cleft that directly appose active zones, and in young adult *lama4*<sup>-/-</sup> mice, the normal precise apposition of active zones to junction folds is lost [20]. Thus laminin 421 may bind to receptors associated with active zones –for example, integrin  $\alpha$ 3 [54]. We speculate that loss of this link leads to decreased stability and eventual disassembly of the nerve terminal.

### Maintenance and Aging of the NMJ

NMJ's react differently to the loss of agrin and laminin- $\alpha$ 4. After agrin is deleted, specializations of the postsynaptic membrane and synaptic cleft are lost; presynaptic alterations occur later, perhaps, as discussed above, as an indirect consequence of the initial disruptions. In the absence of laminin- $\alpha$ 4, in contrast, nerve terminals show gradual structural changes, but the synaptic cleft and postsynaptic membrane are spared. Thus, these two synaptic



**Figure 8. Absence of laminin- $\alpha 4$  causes NMJ to age prematurely.** A. Axon terminals (left) and AChRs (middle panels) from diaphragm muscles of 6 month old *laminin- $\alpha 4$*  mice. The age-associated abnormalities observed in mutants at 6 months of age were similar to those seen in two-year-old wild type mice. B. Prevalence of axonal and terminal changes in *laminin- $\alpha 4$*  and wild type animals. At least 60 NMJs were analyzed for each genotype at each age. Scale Bar = 10  $\mu$ m.

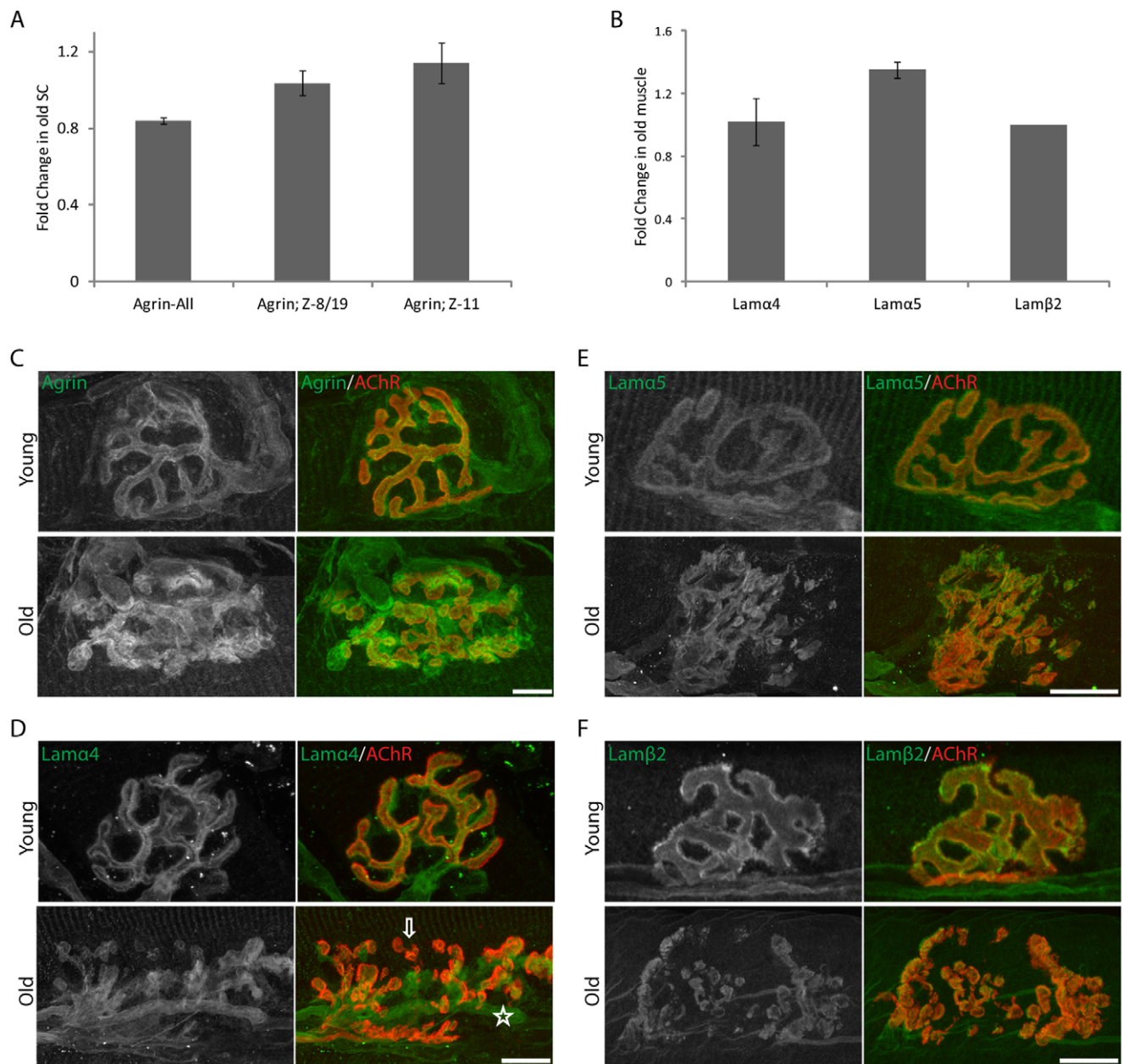
doi:10.1371/journal.pone.0046663.g008

cleft proteins play distinct roles in synaptic maintenance. We draw two main conclusions from these data. First, synaptic organizing molecules, which are critical for embryonic and early postnatal formation of the NMJ, are also required for its long-term maintenance. Second, maintenance of the adult NMJ relies on combined effects of several distinct organizers.

Perhaps our most striking observation is that abnormalities observed in agrin or laminin- $\alpha 4$  mutants resemble those in aged wild-type mice. At present, we have no direct evidence on whether agrin or laminin are involved in the pathogenic mechanisms that underlie synaptic aging. However, the similarity between the mutant phenotypes and normal aging invites the speculation that alterations in these molecules could contribute to age-related synaptic

disassembly. Three types of alterations could occur. First, levels of these components could decline with age. We see no dramatic decline, but our methods would not have allowed us to detect a modest decline. Second, the distribution of the proteins could be altered with age, decreasing their ability to mediate highly localized specializations of appositions. In fact, we have observed such alterations in distribution (Figure 9). Third, the molecular structure of the proteins could change with age. Consistent with this, both neurotrophin and matrix metalloproteinase-3 (MMP-3) cleave agrin, and increased cleavage (or increased levels of cleavage fragments) has been proposed to promote age-related muscle decline, called sarcopenia [55–57]. Likewise, cleavage of laminin  $\alpha$  chains occurs in vivo, and alters the bioactivities of the heterotrimers [58,59]. Such cleavage could





**Figure 9. Molecular changes associated with aging neuromuscular junctions.** A. Levels of RNA encoding different agrin isoforms were compared in the spinal cords of young adult and 2 year old mice by quantitative PCR. B. Levels of laminin- $\alpha$ 4, - $\alpha$ 5 and - $\beta$ 2 RNA were compared in young adult and 2 year old tibialis anterior muscles using quantitative PCR. C–F. Immunostaining with antibodies to agrin and three laminin subunits in young adult and old NMJs. Levels of these proteins did not change significantly with age, but their distribution was altered (see D bottom panel; star = extrasynaptic; arrow = diffuse or lack of expression). Bars represent the SEM. Scale Bar = 10  $\mu$ m. doi:10.1371/journal.pone.0046663.g009

inactive agrin or laminins at the NMJ without affecting the epitopes that our antibodies detect and lead to protein redistribution yet not alter the total levels detectable by staining. A final possibility is that small changes in many different organizers, undetectable by our methods, could lead to age-related synaptic decline.

## Materials and Methods

### Mice

Generation and characterization of agrin null (*agm*<sup>-/-</sup>) [18] agrin conditional (*agm*<sup>flox/flox</sup>) [35], SlickA [36], and laminin  $\alpha$ 4 null

(*lama4*<sup>-/-</sup>) mice [20] have been described previously. To conditionally delete agrin in a YFP marked subset of motor neurons, *agm*<sup>+/-</sup> mice were mated to SLICKA animals to generate heterozygotes and the *agm*<sup>+/-</sup>; *SlickA*<sup>+</sup> offspring were crossed to *agm*<sup>flox/flox</sup> to generate transheterozygotes carrying CreER (*agm*<sup>-/-</sup>; *SlickA*) as well as heterozygote controls (*agm*<sup>+/-</sup>; *SlickA*). Tamoxifen dissolved in corn oil was administered over a two-week period by oral gavage once a day for 4 consecutive days during each week (~3 mg/animal/dose).

For comparative studies of young and old mice, young adult (3 to 4 months) and aged (24 to 28 months) C57BL/6 mice were

obtained from the National Institute on Aging or The Jackson Laboratory (Bar Harbor, Maine) or bred in our vivarium. All experiments conformed to NIH guidelines and were carried out in accordance with protocols approved by the Harvard University Standing Committee on the Use of Animals in Research and Teaching.

### qRT-PCR

For agrin and laminin transcripts analysis, total RNA was prepared from the spinal cords or skeletal muscles of young (3 to 4 months) and old (24 to 28 months) C57BL/6J mice using the RNEasy kit as per the manufacturer's instructions (Qiagen, Valencia, VA). cDNA was generated using the QuantiTect Reverse Transcription system (Qiagen) and transcript levels were determined on a Stratagene MX3000p machine using SYBR green. Primers for the various  $\alpha$  agrin splicing isoforms (total agrin, 8/19, 11) have been described and validated previously [60]. The following primers were used to detect laminin chains:  $\beta$ 2 = GCTTGTAGAGGCCACAGAGG and GACCACTGAGTGCATGGTTG;  $\alpha$ 4 = TGTTTGTGGAGGTGTTCCA and ATGCTCACAGCACCAGTAC;  $\alpha$ 5 = TACAACCTGTGACCCGACAA and GGATGACAAGAGCCAGCTTC.

### Immunohistochemistry

Mice were anesthetized with sodium pentobarbital and then transcardially perfused with phosphate buffered saline (PBS) followed by 4% paraformaldehyde (PFA) in PBS. Muscles were postfixed in PFA for 10 min and used for whole mount analysis.

For agrin staining, muscles were first exposed to ice cold methanol (100%) for 10 s and then washed in PBS. Tissue was then incubated successively with blocking solution (5% normal goat serum, 3% BSA and 0.5% Triton-X-100 in PBS) overnight at 4°C, primary antibodies diluted in blocking solution for 12–14 hours at 4°C, and fluorophore-labeled secondary antibodies or

fBTX (Molecular Probes, Eugene, OR) for 2 to 3 hours at room temperature.

The primary antibodies used in this study were directed against the following molecules: agrin (1:500, gift from Z. Hall); laminin- $\alpha$ 4 and laminin- $\beta$ 2 (1:1000, gifts from T. Sasaki, Shriners Research Center, Portland OR); laminin- $\alpha$ 5 (1:1000, [61]); rapsyn (1:250, Thermo),  $\beta$ -dystroglycan (1:250, Developmental Studies Hybridoma Bank), GFP (1:1000, Abcam); neurofilament (SMI312, 1:1000, Covance); and syntrophin 2 (Synt2, 1:250, Developmental Studies Hybridoma Bank). Fluorescently labeled BTX (Alexa 488, 594, or 647; Invitrogen) was used to mark AChRs.

### Histological Analysis

Maximum intensity projections of confocal stacks were generated using Image J or Metamorph 6.03 software (Molecular Devices, Sunnyvale, CA). For structural analysis in *lama4*<sup>-/-</sup> mice, sprouting was defined as an axonal profile that extended beyond  $\geq 2$   $\mu$ m of the endplate; partial or complete denervation was assayed by the apposition of the axon terminals and their AChRs; multiple innervation was identified by counting the number of axons impinging onto each endplate. For agrin conditional animals, altered NMJs were quantified as those that displayed any one of the several dystrophies outlined above, including axon swelling, nerve sprouting, atrophy, fragmentation, and thinning. For axon terminal pair analyses, whole triangularis muscles from conditional agrin mice were imaged and reconstructed. Images were scanned to identify those in which two terminals arose from the same axon and their level of dystrophy was then evaluated relative to control animals.

### Author Contributions

Conceived and designed the experiments: MAS GV JCT JWL JRS. Performed the experiments: MAS GV JCT. Analyzed the data: MAS GV JCT. Wrote the paper: MAS GV JRS.

### References

- Lichtman JW, Magrassi L, Purves D (1987) Visualization of neuromuscular junctions over periods of several months in living mice. *J Neurosci* 7: 1215–1222.
- Wigston DJ (1990) Repeated in vivo visualization of neuromuscular junctions in adult mouse lateral gastrocnemius. *J Neurosci* 10: 1753–1761.
- Balicer-Gordon RJ, Lichtman JW (1990) In vivo visualization of the growth of pre- and postsynaptic elements of neuromuscular junctions in the mouse. *J Neurosci* 10: 894–908.
- Balicer-Gordon RJ (1997) Age-related changes in neuromuscular innervation. *Muscle Nerve Suppl* 5: S83–87.
- Cardasis CA, LaFontaine DM (1987) Aging rat neuromuscular junctions: a morphometric study of cholinesterase-stained whole mounts and ultrastructure. *Muscle Nerve* 10: 200–213.
- Fahim MA, Robbins N (1982) Ultrastructural studies of young and old mouse neuromuscular junctions. *J Neurocytol* 11: 641–656.
- Fahim MA, Holley JA, Robbins N (1983) Scanning and light microscopic study of age changes at a neuromuscular junction in the mouse. *J Neurocytol* 12: 13–25.
- Valdez G, Tapia JC, Kang H, Clemenson GD Jr, Gage FH, et al. (2010) Attenuation of age-related changes in mouse neuromuscular synapses by caloric restriction and exercise. *Proc Natl Acad Sci U S A* 107: 14863–14868.
- Valdez G, Tapia JC, Lichtman JW, Fox MA, Sanes JR (2012) Shared resistance to aging and ALS in neuromuscular junctions of specific muscles. *PLoS One* 7: e34640. doi: 10.1371/journal.pone.0034640.
- Fischer LR, Culver DG, Tennant P, Davis AA, Wang M, et al. (2004) Amyotrophic lateral sclerosis is a distal axonopathy: evidence in mice and man. *Exp Neurol* 185: 232–240.
- Gurney ME, Pu H, Chiu AY, Dal Canto MC, Polchow CY, et al. (1994) Motor neuron degeneration in mice that express a human Cu,Zn superoxide dismutase mutation. *Science* 264: 1772–1775.
- Gomez AM, Van Den Broeck J, Vrolix K, Janssen SP, Lemmens MA, et al. (2010) Antibody effector mechanisms in myasthenia gravis-pathogenesis at the neuromuscular junction. *Autoimmunity* 43: 353–370.
- Schaefer AM, Sanes JR, Lichtman JW (2005) A compensatory subpopulation of motor neurons in a mouse model of amyotrophic lateral sclerosis. *J Comp Neurol* 490: 209–219.
- Sanes JR, Lichtman JW (2001) Induction, assembly, maturation and maintenance of a postsynaptic apparatus. *Nat Rev Neurosci* 2: 791–805.
- Kummer TT, Misgeld T, Sanes JR (2006) Assembly of the postsynaptic membrane at the neuromuscular junction: paradigm lost. *Curr Opin Neurobiol* 16: 74–82.
- Wu H, Xiong WC, Mei L (2010) To build a synapse: signaling pathways in neuromuscular junction assembly. *Development* 137: 1017–1033.
- Noakes PG, Gautam M, Mudd J, Sanes JR, Merlie JP (1995) Aberrant differentiation of neuromuscular junctions in mice lacking  $\alpha$ -laminin/laminin beta 2. *Nature* 374: 258–262.
- Gautam M, Noakes PG, Moscoso L, Rupp F, Scheller RH, et al. (1996) Defective neuromuscular synaptogenesis in agrin-deficient mutant mice. *Cell* 85: 525–535.
- Burgess RW, Nguyen QT, Son YJ, Lichtman JW, Sanes JR (1999) Alternatively spliced isoforms of nerve- and muscle-derived agrin: their roles at the neuromuscular junction. *Neuron* 23: 33–44.
- Patton BL, Cunningham JM, Thyboll J, Kortessma J, Westerblad H, et al. (2001) Properly formed but improperly localized synaptic specializations in the absence of laminin alpha4. *Nat Neurosci* 4: 597–604.
- Fox MA, Sanes JR, Borza DB, Eswarakumar VP, Fessler R, et al. (2007) Distinct target-derived signals organize formation, maturation, and maintenance of motor nerve terminals. *Cell* 129: 179–193.
- DeChiara TM, Bowen DC, Valenzuela DM, Simmons MV, Poueymirou WT, et al. (1996) The receptor tyrosine kinase MuSK is required for neuromuscular junction formation in vivo. *Cell* 85: 501–512.
- Glass DJ, Bowen DC, Stitt TN, Radziejewski C, Bruno J, et al. (1996) Agrin acts via a MuSK receptor complex. *Cell* 85: 513–523.
- Kim N, Stiegler AL, Cameron TO, Hallock PT, Gomez AM, et al. (2008) Lrp4 is a receptor for Agrin and forms a complex with MuSK. *Cell* 135: 334–342.
- Zhang B, Luo S, Wang Q, Suzuki T, Xiong WC, et al. (2008) LRP4 serves as a coreceptor of agrin. *Neuron* 60: 285–297.
- Yang X, Arber S, William C, Li L, Tanabe Y, et al. (2001) Patterning of muscle acetylcholine receptor gene expression in the absence of motor innervation. *Neuron* 30: 399–410.



27. Lin W, Burgess RW, Dominguez B, Pfaff SL, Sanes JR, et al. (2001) Distinct roles of nerve and muscle in postsynaptic differentiation of the neuromuscular synapse. *Nature* 410: 1057–1064.
28. Lin W, Dominguez B, Yang J, Aryal P, Brandon EP, et al. (2005) Neurotransmitter acetylcholine negatively regulates neuromuscular synapse formation by a Cdk5-dependent mechanism. *Neuron* 46: 569–579.
29. Misgeld T, Kummer TT, Lichtman JW, Sanes JR (2005) Agrin promotes synaptic differentiation by counteracting an inhibitory effect of neurotransmitter. *Proc Natl Acad Sci U S A* 102: 11088–11093.
30. Flanagan-Steet H, Fox MA, Meyer D, Sanes JR (2005) Neuromuscular synapses can form in vivo by incorporation of initially aneural postsynaptic specializations. *Development* 132: 4471–4481.
31. Miner JH, Yurchenco PD (2004) Laminin functions in tissue morphogenesis. *Annu Rev Cell Dev Biol* 20: 255–284.
32. Patton BL, Miner JH, Chiu AY, Sanes JR (1997) Distribution and function of laminins in the neuromuscular system of developing, adult, and mutant mice. *J Cell Biol* 139: 1507–1521.
33. Patton BL, Chiu AY, Sanes JR (1998) Synaptic laminin prevents glial entry into the synaptic cleft. *Nature* 393: 698–701.
34. Nishimune H, Valdez G, Jarad G, Moulson CL, Muller U, et al. (2008) Laminins promote postsynaptic maturation by an autocrine mechanism at the neuromuscular junction. *J Cell Biol* 182: 1201–1215.
35. Harvey SJ, Jarad G, Cunningham J, Rops AL, van der Vlag J, et al. (2007) Disruption of glomerular basement membrane charge through podocyte-specific mutation of agrin does not alter glomerular permselectivity. *Am J Pathol* 171: 139–152.
36. Young P, Qiu L, Wang D, Zhao S, Gross J, et al. (2008) Single-neuron labeling with inducible Cre-mediated knockout in transgenic mice. *Nat Neurosci* 11: 721–728.
37. Feng G, Mellor RH, Bernstein M, Keller-Peck C, Nguyen QT, et al. (2000) Imaging neuronal subsets in transgenic mice expressing multiple spectral variants of GFP. *Neuron* 28: 41–51.
38. Hoch W, Ferns M, Campanelli JT, Hall ZW, Scheller RH (1993) Developmental regulation of highly active alternatively spliced forms of agrin. *Neuron* 11: 479–490.
39. Sanes JR (2003) The basement membrane/basal lamina of skeletal muscle. *J Biol Chem* 278: 12601–12604.
40. Balice-Gordon RJ, Breedlove SM, Bernstein S, Lichtman JW (1990) Neuromuscular junctions shrink and expand as muscle fiber size is manipulated: in vivo observations in the androgen-sensitive bulbocavernosus muscle of mice. *J Neurosci* 10: 2660–2671.
41. Brown MC, Holland RL, Hopkins WG (1981) Motor nerve sprouting. *Annu Rev Neurosci* 4: 17–42.
42. Andonian MH, Fahim MA (1988) Endurance exercise alters the morphology of fast- and slow-twitch rat neuromuscular junctions. *Int J Sports Med* 9: 218–223.
43. Deschenes MR, Tenny KA, Wilson MH (2006) Increased and decreased activity elicits specific morphological adaptations of the neuromuscular junction. *Neuroscience* 137: 1277–1283.
44. Sanes JR, Feldman DH, Cheney JM, Lawrence JC Jr (1984) Brain extract induces synaptic characteristics in the basal lamina of cultured myotubes. *J Neurosci* 4: 464–473.
45. Kim N, Burden SJ (2008) MuSK controls where motor axons grow and form synapses. *Nat Neurosci* 11: 19–27.
46. Yumoto N, Kim N, Burden SJ (2012) Lrp4 is a retrograde signal for presynaptic differentiation at neuromuscular synapses. *Nature* doi:10.1038/nature11348.
47. Marangi PA, Forsayeth JR, Mittaud P, Erb-Vogtli S, Blake DJ, et al. (2001) Acetylcholine receptors are required for agrin-induced clustering of postsynaptic proteins. *EMBO J* 20: 7060–7073.
48. Campagna JA, Ruegg MA, Bixby JL (1997) Evidence that agrin directly influences presynaptic differentiation at neuromuscular junctions in vitro. *Eur J Neurosci* 9: 2269–2283.
49. Burgess RW, Dickman DK, Nunez L, Glass DJ, Sanes JR (2002) Mapping sites responsible for interactions of agrin with neurons. *J Neurochem* 83: 271–284.
50. Wu H, Lu Y, Shen C, Patel N, Gan L, et al. (2012) Distinct Roles of Muscle and Motoneuron LRP4 in Neuromuscular Junction Formation. *Neuron* 75: 94–107.
51. Hesser BA, Henschel O, Witzemann V (2006) Synapse disassembly and formation of new synapses in postnatal muscle upon conditional inactivation of MuSK. *Mol Cell Neurosci* 31: 470–480.
52. Bezakova G, Helm JP, Francolini M, Lomo T (2001) Effects of purified recombinant neural and muscle agrin on skeletal muscle fibers in vivo. *J Cell Biol* 153: 1441–1452.
53. Miyamaru S, Kumai Y, Ito T, Yumoto E (2008) Effects of long-term denervation on the rat thyroarytenoid muscle. *Laryngoscope* 118: 1318–1323.
54. Cohen MW, Hoffstrom BG, DeSimone DW (2000) Active zones on motor nerve terminals contain alpha 3beta 1 integrin. *J Neurosci* 20: 4912–4921.
55. Bolliger MF, Zurlinden A, Luscher D, Butikofer L, Shakhova O, et al. (2010) Specific proteolytic cleavage of agrin regulates maturation of the neuromuscular junction. *J Cell Sci* 123: 3944–3955.
56. Butikofer L, Zurlinden A, Bolliger MF, Kunz B, Sonderegger P (2011) Destabilization of the neuromuscular junction by proteolytic cleavage of agrin results in precocious sarcopenia. *FASEB J* 25: 4378–4393.
57. Werle MJ, VanSaun M (2003) Activity dependent removal of agrin from synaptic basal lamina by matrix metalloproteinase 3. *J Neurocytol* 32: 905–913.
58. Bair EL, Chen ML, McDaniel K, Sekiguchi K, Cress AE, et al. (2005) Membrane type 1 matrix metalloprotease cleaves laminin-10 and promotes prostate cancer cell migration. *Neoplasia* 7: 380–389.
59. Baudoin C, Fantin L, Meneguzzi G (2005) Proteolytic processing of the laminin alpha3 G domain mediates assembly of hemidesmosomes but has no role on keratinocyte migration. *J Invest Dermatol* 125: 883–888.
60. Ruggiu M, Herbst R, Kim N, Jevsek M, Fak JJ, et al. (2009) Rescuing Z+ agrin splicing in Nova null mice restores synapse formation and unmasks a physiologic defect in motor neuron firing. *Proc Natl Acad Sci U S A* 106: 3513–8.
61. Miner JH, Patton BL, Lentz SI, Gilbert DJ, Snider WD, et al. (1997) The laminin alpha chains: expression, developmental transitions, and chromosomal locations of alpha1–5, identification of heterotrimeric laminins 8–11, and cloning of a novel alpha3 isoform. *J Cell Biol* 137: 685–701.